

Next-Generation Topical Delivery for Infantile Dermatitis: Exploring the Therapeutic Potential of Solid Lipid Nanoparticles

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ABSTRACT

Background. Infantile dermatitis, most commonly atopic in origin, affects an estimated 15–20% of children worldwide and imposes a substantial burden on quality of life and household finances. The infant skin barrier is structurally and functionally immature: the stratum corneum is roughly 30% thinner than in adults, transepidermal water loss is higher, and the body-surface-area-to-mass ratio is approximately 2.5–3 times greater. These features raise the risk of systemic exposure from topical therapy precisely in the population least able to tolerate it, and they constrain the safe use of the current standard of care.

Scope. This narrative review examines solid lipid nanoparticles (SLNs) and their second-generation counterparts, nanostructured lipid carriers (NLCs), as a topical delivery platform for infantile dermatitis. It covers carrier architecture and composition, production and characterization methodology, mechanisms of cutaneous delivery, the published formulation evidence for dermatitis-relevant actives, and the safety, regulatory and manufacturing considerations governing translation.

Key findings. SLNs deliver drug to the skin through three interacting mechanisms: formation of an occlusive lipid film that reduces transepidermal water loss and hydrates the stratum corneum, size-dependent surface contact and follicular deposition, and sustained release from a solid lipid matrix. Across the formulations reviewed — fluticasone propionate, tacrolimus, ebastine, etofenamate, ibuprofen, curcumin, resveratrol, quercetin and leflunomide — encapsulation consistently improved physical and photochemical stability and moderated the release profile relative to free drug. Where studies progressed to animal disease models, comparable or superior anti-inflammatory efficacy was achieved at substantially reduced doses, including an 8- to 16-fold dose reduction for etofenamate and ibuprofen.

Conclusion. The mechanistic case for SLN-based topical therapy in infantile dermatitis is coherent and is supported by consistent adult and adult-animal data. It is not, however, supported by pediatric evidence. No permeation study identified used infant or neonatal skin, no in vivo study used a neonatal or juvenile animal model, no clinical trial of an SLN formulation for infantile dermatitis was identified, and no regulatory authority has approved such a product. Until pediatric-specific pharmacokinetic, irritation and safety data are generated, SLN delivery for infantile dermatitis should be regarded as a mechanistically promising but clinically unproven strategy.

Keywords: solid lipid nanoparticles; nanostructured lipid carriers; infantile atopic dermatitis; topical drug delivery; pediatric formulation; skin barrier

1. INTRODUCTION

1.1 Disease burden and epidemiology

Atopic dermatitis (AD) is a chronic, relapsing inflammatory skin disorder that often travels alongside asthma, allergic rhinitis and food allergy ¹. It is common - an estimated 15–20% of children and 1–3% of adults are affected worldwide - and its burden reaches beyond the skin itself: pruritus disrupts sleep in nearly all patients, and families absorb the cost in physician visits, medication and missed school or work ¹.

Reported cumulative incidence ranges from 11% to 21% depending on age group and region ². In a clinicoepidemiological study from northern India, infants with AD had a mean onset of 4.2 months and a disease duration of 3.3 months, against a 4.1-year mean onset and 1.9-year duration in older children, with urban children over-represented ³. A hospital-based study from the same region found AD to be the single commonest skin condition among pediatric dermatology attendees (29.9% of cases; male-to-female ratio 2.25:1; mean onset 4.5 months), with infants presenting mainly as facial, acute eczema and older children showing more chronic, variable disease; symptoms worsened in winter for 62% of patients ⁴. A much lower prevalence, 0.42%, was reported among dermatology outpatients in eastern India ⁵ - a spread that reflects how much regional methodology, not just regional biology, shapes these numbers.

Only three drug classes carry FDA approval for topical AD therapy: corticosteroids, two calcineurin inhibitors, and one phosphodiesterase-4 (PDE4) inhibitor. Because AD is usually lifelong, the treatment needs to be both effective and tolerable long-term. Topical corticosteroids (TCS) remain first-line, but sustained use brings real risks - thinning skin, telangiectasia, striae, and occasionally enough systemic absorption to affect the hypothalamic-pituitary-adrenal axis ⁶⁻⁷ - and these risks are not abstract to caregivers: steroid apprehension is well documented and it undermines adherence ⁸. Topical calcineurin inhibitors (pimecrolimus, tacrolimus) have been marketed since 2000; a 2006 boxed warning over a theoretical malignancy risk has not been borne out by later research ⁹. Crisaborole, the PDE4 inhibitor, was approved in 2016 and has shown an encouraging early safety record.

1.2 Infant skin physiology relative to adult skin

Infant skin is not simply a smaller version of adult skin - it differs across structural, biochemical and functional parameters, several of which bear directly on how a topical formulation behaves once applied. The stratum corneum and overall epidermis are thinner, corneocytes are smaller, natural moisturizing factor and water-holding capacity are reduced, and transepidermal water loss is elevated at birth and falls only gradually thereafter. Melanocytes and Langerhans cells remain functionally immature, and surface pH runs higher. Together, these features leave a barrier that is more permeable, more easily irritated and less able to hold water than its adult counterpart (Table 1).

Table 1. Structural and functional characteristics of healthy infant skin compared with adult skin.

Parameter	Healthy infant skin compared with adult skin
Epidermal cell size	Smaller corneocytes and keratinocytes ¹⁰
Epidermal surface	Denser network of surface microrelief ¹⁰
Epidermal thickness	Stratum corneum approximately 30% thinner; overall epidermis approximately 20% thinner ¹⁰
Dermal organization	More uniform dermal papillae; extensive but less organized vascular network; lower density of collagen fibre bundles ¹⁰⁻¹²
Natural moisturizing factor (NMF)	Lower ¹³
Melanin	Lower ¹⁴
Water content	Lower at birth, rising gradually over the first year ¹⁰
Cell turnover	Higher ¹⁰
Hydration and water-holding capacity	Lower hydration at birth, peaking between 3-12 months; water-holding capacity remains lower ^{10,13}
Immunological barrier	Epidermal Langerhans cells not yet fully matured ¹⁴
Skin pH	Higher ¹⁰
Photoprotection	Melanocytes not yet fully matured ¹⁴
Sebaceous activity	Higher at birth, dropping sharply within the first few days ¹⁵
Transepidermal water loss (TEWL)	Higher at birth, decreasing gradually over the first few years ¹⁴

1.3 Current standard of care and its limitations in infants

Topical corticosteroids (TCS) remain the anti-inflammatory cornerstone of AD treatment, particularly for flares, once basic skin care and moisturizing alone prove insufficient. Repeated courses of low- to mid-potency TCS applied until clearance are supported by RCT evidence extending to five years in children and one year in adults¹⁶, and a proactive strategy - twice-weekly application to stabilized skin to pre-empt flares - has outperformed reactive treatment alone in a year-long pediatric RCT¹⁷⁻¹⁸. The British National Formulary sorts TCS into four potency tiers¹⁹, and judicious use of low- to mid-potency agents carries an acceptable long-term safety profile. Higher-potency formulations used over extended periods, however, can cause striae, atrophy, telangiectasia, burning and acneiform eruptions, and rare systemic effects such as HPA-axis suppression occur more often in children, whose body-surface-to-mass ratio runs roughly 2.5 to 3 times that of adults²⁰. Twice-weekly proactive dosing has not itself been linked to atrophy¹⁷, though no consensus exists on how much TCS to apply per use.

Topical calcineurin inhibitors (TCIs) - tacrolimus and pimecrolimus - have been approved for AD since 2002, with RCT support for twice-daily intermittent use from three months of age and follow-up extending to five years²¹. A meta-analysis of 25 RCTs found 0.1% tacrolimus comparable to mid-potency hydrocortisone butyrate 0.1%, while 0.03% tacrolimus underperformed that comparator but beat low-potency hydrocortisone acetate 1%²². Approved ages differ by product (pimecrolimus 1% and tacrolimus 0.03% from age 2; tacrolimus 0.1% from age 16), and proactive tacrolimus use has shown year-long effectiveness and safety in both children and adults. The most common adverse effect, a transient burning sensation, resolves within a week in roughly 80% of patients; TCIs do not raise bacterial infection risk, though herpes simplex risk is modestly elevated²³. The 2006 malignancy boxed warning has not been substantiated by long-term controlled trials or registry data, including the largest and longest pediatric pimecrolimus study to date²⁴.

Emollients sit at the centre of AD management because xerosis and barrier impairment define the disease. Regular moisturizer use repairs barrier structure, cuts transepidermal water loss and restores the lipid barrier's water-handling capacity²⁵, with RCT evidence for both short- and long-term steroid-sparing effects and flare prevention²⁶. No trial has fixed an optimal dose, but guidance generally calls for at least twice-daily application - roughly 250 g weekly in adults²⁷ - applied within three minutes of bathing and continued even once AD has cleared. Newer prescription emollient devices have shown no clear advantage over over-the-counter moisturizers in head-to-head trials²⁸. Some ingredients carry their own anti-inflammatory activity: an aloe vera-olive oil cream matched 0.1% betamethasone for chronic sulfur-mustard-exposure lesions over six weeks²⁹, virgin coconut oil showed moderate activity in a rat ear-oedema model³⁰, and shea butter's triterpene esters have shown anti-inflammatory and antitumor effects in vivo and in vitro³¹. In practice, the most reliable way to limit steroid exposure combines consistent baseline emollient care with early anti-inflammatory treatment to head off flares before they escalate³².

Why this matters more in infants. Four constraints converge in this population: a proportionally larger body surface area relative to mass raises the risk of meaningful systemic corticosteroid absorption; extended or improper potent-steroid use can atrophy skin that is already thinner; sustained anti-inflammatory therapy needs a level of clinical oversight that is not always available; and frequent reapplication, compounded by caregiver anxiety over steroid side effects, undermines adherence. A delivery system that concentrates drug at the application site, sustains local availability and limits the fraction reaching systemic circulation would address all four at once.

1.4 Rationale for lipid nanoparticle-based delivery

Skin is the body's primary barrier against chemical, mechanical, physical and microbial insult, yet it remains vulnerable to rashes, infection, burns, wounds and tumours³³⁻³⁹. Topical delivery treats disease directly at the site and can limit systemic exposure, but its effectiveness hinges on the vehicle's ability to actually get drug into the skin. Conventional creams, gels, ointments, pastes and oils are widely used but limited by poor penetration, local irritation and weak drug retention, which has pushed nanotechnology-based delivery - designed to improve penetration, stability and controlled release - to the forefront of skin-disorder therapeutics⁴⁰⁻⁴¹.

Among the lipid-based nanosystems explored for topical use - liposomes, ethosomes, transferosomes, SLNs, NLCs, cubosomes and monoolein dispersions - biocompatibility and lipid composition allow efficient loading of lipophilic drugs, controlled release, improved bioavailability and fewer adverse effects⁴²⁻⁴³. Their compositional resemblance to native stratum corneum lipids promotes skin interaction, hydration and potentially deeper penetration.

SLNs and NLCs stand out for dermal use because of their solid lipid matrix and controlled-release capacity: SLNs are built mainly from solid lipids, while NLCs blend solid and liquid lipids to create structural imperfections that raise drug-loading capacity. Both are typically made from biocompatible, GRAS lipids, and both can protect drugs from degradation while improving skin adhesion and penetration. Their small size also produces an occlusive film that lowers transepidermal water loss and raises stratum corneum hydration - potentially enhancing permeation

while independently treating barrier-compromised skin. In an infant with dermatitis, where the barrier is already impaired and the safety margin is narrow, that combination is not incidental - it targets the specific pharmacological problem this population presents.

1.5 Objective and scope

This review builds the case for, and weighs the evidence gaps against, SLN-based topical therapy in infantile dermatitis, moving through carrier architecture and production (Section 3), the mechanisms of cutaneous delivery and their relevance to an immature barrier (Section 4), the published formulation evidence for dermatitis-relevant actives (Section 5), and the toxicological, regulatory and manufacturing picture (Section 6). Throughout, it draws a clear line between what has been shown in adults and adult animal models and what has been shown in infants, since that boundary - not any mechanistic doubt - is the main obstacle to clinical translation.

2. MATERIALS AND METHODS

Literature was retrieved from PubMed, Scopus, ScienceDirect and Google Scholar, with targeted searches of ClinicalTrials.gov, Google Patents and WIPO PATENTSCOPE for trial and intellectual-property status, and of the FDA, EMA and ICH repositories for regulatory guidance. The search covered January 2000 to the date of submission, prioritizing work from the preceding seven years while retaining older sources that describe foundational methodology still in use.

Search terms combined four clusters: carrier terms (“solid lipid nanoparticles”, “SLN”, “nanostructured lipid carriers”, “NLC”, “lipid nanoparticles”); route terms (“topical”, “dermal”, “transdermal”, “skin permeation”, “follicular delivery”); disease terms (“atopic dermatitis”, “eczema”, “allergic contact dermatitis”, “infantile dermatitis”, “pediatric dermatology”); and active-agent terms (“corticosteroid”, “tacrolimus”, “pimecrolimus”, “antihistamine”, “curcumin”, “quercetin”). Reference lists of retrieved reviews were hand-searched for additional primary studies.

A study was carried forward for the Section 5 evidence base if it described an SLN or NLC system loaded with a drug relevant to inflammatory skin disease - corticosteroids, calcineurin inhibitors, antihistamines, NSAIDs or plant-derived anti-inflammatory actives - and reported at least basic physicochemical characterization alongside a permeation, release or efficacy endpoint. Studies confined to non-inflammatory indications (isolated antifungal or oncology applications, for instance) were excluded unless they offered a directly useful methodological comparison, and only English-language publications were included. This is a narrative rather than a systematic review: no formal risk-of-bias assessment or quantitative synthesis was undertaken, and the search was not registered.

3. RESULTS AND DISCUSSION

This section presents and discusses the findings drawn from the literature review, moving through carrier technology and composition, mechanisms of cutaneous delivery, the formulation evidence base, and the safety, regulatory and translational picture.

Solid lipid nanoparticles are colloidal drug carriers built from a lipid matrix that stays solid at both room and body temperature, dispersed in an aqueous medium and stabilized by one or more surfactants. First introduced in the early 1990s as an alternative to liposomes, polymeric nanoparticles and conventional emulsions, SLNs occupy a submicron size range generally cited as roughly 50–1000 nm⁴⁴. The matrix draws on physiological, biodegradable, biocompatible lipids that accommodate lipophilic and, to a lesser extent, hydrophilic actives - a property well suited to the anti-inflammatory and antipruritic agents relevant to infantile dermatitis⁴⁴. This section covers the structural classification of lipid nanoparticles, their building blocks, the main production routes and the physicochemical parameters used to characterize them.

3.1 Definition and structural concept

Structurally, an SLN can be pictured as an oil-in-water nanoemulsion in which the dispersed lipid droplet has been replaced by a solid lipid core - the solid state at physiological temperature being what separates SLNs from nanoemulsions, whose dispersed phase stays liquid. That rigidity governs how drug is retained and released: depending on lipid composition, drug-lipid interactions and processing conditions, the drug may dissolve within the matrix or sit closer to the particle surface, and these structural choices in turn set drug loading, entrapment efficiency, particle size, release kinetics and physical stability⁴⁴.

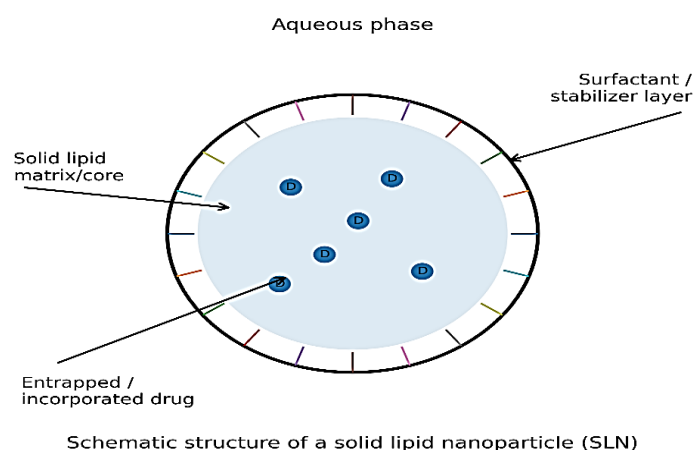


Figure 1. Schematic structure of a solid lipid nanoparticle (SLN).

3.2 First-generation SLN versus second-generation NLC

SLNs are conventionally treated as the first generation of solid lipid carrier systems at the nanometre scale ⁴⁵. Relative to earlier colloidal systems, they offer controlled or sustained release, protect chemically unstable molecules such as retinoids and peptides from degradation, rely on relatively low-cost and pharmaceutically acceptable excipients, and work across oral, parenteral, ocular, pulmonary and dermal routes ⁴⁵. The same highly ordered crystalline lattice that gives them this stability, though, brings two well-known limitations: drug loading is often low, and drug can be expelled during storage as the matrix shifts from the less-ordered α form toward the tightly packed, thermodynamically stable β form generated during cooling and crystallization ^{44,46}.

Nanostructured lipid carriers were developed as the second generation specifically to address these gaps ⁴⁴⁻⁴⁵. The NLC matrix blends a solid lipid with a liquid lipid (oil) at a ratio that keeps the mixture solid at room and body temperature; the liquid lipid disrupts the regular crystal lattice, opening structural irregularities that accommodate more drug and amorphous clusters. NLCs generally achieve higher loading, less storage-related drug expulsion, and better long-term physical stability than conventional SLNs as a result ^{44,47}, and fall into three recognised subtypes: imperfect-type, where a moderate amount of liquid lipid creates lattice defects while the particle largely retains its solid character; amorphous-type, where the lipid blend is chosen to solidify without crystallizing and so avoid the polymorphic shifts that expel drug; and multiple-type (oil-in-solid-lipid-in-water), where small oil compartments dispersed through the solid matrix - analogous to a w/o/w emulsion - allow higher loading of very lipophilic actives while the surrounding shell still confers controlled release.

For topical delivery, the choice between SLN and NLC comes down to the drug's physicochemical properties and the required loading capacity, release profile and stability, with NLCs generally favoured where higher loading and reduced storage expulsion matter most ^{44-45,47}.

3.3 Composition

Solid lipids form the structural backbone and are typically drawn from four categories: fatty acids (stearic, palmitic acid); mono-, di- and triglycerides and complex glyceride mixtures (glyceryl monostearate, glyceryl behenate/Compritol® 888 ATO, glyceryl palmitostearate/Precirol® ATO 5, tripalmitin, trilaurin); waxes (cetyl palmitate, beeswax); and steroids (cholesterol) ^{44,48}. Chain length, branching and lipid purity influence matrix crystallinity, which in turn affects loading, entrapment efficiency and the likelihood of a polymorphic shift during storage.

Liquid lipids for NLC formulation - commonly medium-chain triglycerides, oleic acid, isopropyl myristate, or vegetable oils such as almond, sunflower or coconut oil - are blended with the solid lipid to disrupt its crystalline structure, opening space for more drug while the matrix stays solid at room and physiological temperature ⁴⁴⁻⁴⁵.

Surfactants and stabilizers adsorb at the lipid-water interface during production, lowering interfacial tension and stabilizing the particles electrostatically and/or sterically. Non-ionic surfactants are generally preferred for their lower irritancy - a consideration that matters more for an infant-facing product. Poloxamer 188, a triblock PEO-PPO-PEO copolymer, is widely used for steric stabilization ⁴⁹, while polysorbates (Tween 20/80) and sorbitan esters (Span series) are chosen by hydrophilic-lipophilic balance ^{48,50}. PEGylated stabilizers such as macrogol-15-hydroxystearate add steric stabilization, and phospholipids like soy or egg lecithin can serve as co-surfactants that improve biological-membrane compatibility ⁴⁸. Ionic surfactants (sodium cholate, sodium deoxycholate, sodium dodecyl sulfate) improve emulsification and modulate surface charge but carry a higher

irritation risk, so surfactant choice should track both the administration route and the formulation's specific requirements^{48,50}.

3.4 Production methods

SLNs and NLCs can be manufactured by high-energy (mechanical) and low-energy (emulsification- or solvent-based) techniques. The four methods most widely reported for dermatological development are compared in Table 2⁵¹⁻⁵³.

Table 2. Comparison of the principal production methods used for SLNs/NLCs.

Method	Principle	Advantages	Limitations
Hot high-pressure homogenization (HPH)	Drug-loaded lipid melt is pre-emulsified, then forced through a narrow homogenizing gap (100-2000 bar); recrystallizes on cooling	Solvent-free; fast; scalable to pilot/industrial batches; narrow, reproducible size distribution	Thermal/shear stress unsuitable for heat-labile actives; possible drug loss to the aqueous phase; aggregation on cooling/storage
Microemulsion dilution	A warm, transparent O/W microemulsion is diluted into excess cold water (2-3 degC); particles precipitate instantly	Low mechanical energy; no specialized equipment; mild conditions suit labile drugs	High surfactant load; dilute product needs downstream concentration; batch reproducibility and scale-up are hard
Solvent emulsification-diffusion/evaporation	Lipid dissolved in a partly water-miscible solvent, emulsified into aqueous surfactant, then solvent removed	Room-temperature process; suits thermolabile and peptide actives	Residual solvent is a regulatory concern, especially for infant products; lower loading; slower processing
Ultrasonication/high-shear homogenization	Lipid melt dispersed by rotor-stator mixing, then probe-sonicated before cooling	Simple, low-cost, accessible bench-scale equipment	Broad size distribution; possible metal contamination from probe erosion; poor reproducibility

Beyond these, emerging techniques - microfluidic mixing, phase-inversion temperature processing, membrane-contactor methods and supercritical-fluid production - offer finer control over particle characteristics and can cut or eliminate organic solvent use, though their suitability still depends on formulation, process requirements, scalability and the drug involved⁵¹. For a pediatric topical formulation, method choice should weigh residual solvents, reproducibility, scalability and API stability, with solvent limits and excipient burden carrying extra weight relative to an adult product.

3.5 Characterization parameters

A consistent panel of characterization parameters is essential for formulation optimization and batch-to-batch reproducibility. Table 3 summarizes the parameters commonly reported in the SLN/NLC literature, the techniques used to determine them, and the typical ranges reported for dermal formulations⁵⁴⁻⁵⁵.

Table 3. Core physicochemical characterization parameters for SLNs/NLCs.

Parameter	Technique	What it shows	Typical dermal range
Particle size and PDI	Dynamic light scattering	Mean hydrodynamic diameter and size homogeneity; influences skin deposition and follicular access	~70-300 nm; PDI < 0.3 generally acceptable
Zeta potential	Electrophoretic light scattering	Net surface charge and indicator of colloidal stability (sterically stabilized systems can remain stable at lower absolute values)	Typically -10 to -45 mV, lipid/surfactant-dependent
Entrapment efficiency (%EE)	Ultracentrifugation/ultrafiltration + UV or HPLC assay of free drug	Proportion of drug entrapped within the particles	~60% to >95%, depending on drug lipophilicity and lipid crystallinity
Crystallinity/polymorphism	Differential scanning calorimetry (DSC)	Thermal behaviour, degree of crystallinity, polymorphic form, drug-lipid interactions	Melting endotherm often depressed and broadened relative to bulk lipid
Crystal structure	X-ray diffraction (XRD/PXRD)	Lattice structure; attenuated drug peaks can indicate amorphization	Attenuation/disappearance of sharp drug diffraction peaks
Morphology	TEM/SEM/AFM	Particle shape and surface texture; complements DLS sizing	Predominantly spherical to near-spherical, smooth-surfaced

Particle size and PDI shape drug release, skin deposition and follicular localization: smaller, more homogeneous nanoparticles achieve closer surface contact and may favour follicular deposition, though formulation and skin condition both modulate the effect ⁵⁶. Zeta potential indicates colloidal stability, but its reading depends on whether stabilization is mainly electrostatic or steric ⁴⁹. DSC and XRD/PXRD are complementary solid-state techniques - the former tracking thermal behaviour and crystallinity, the latter crystal structure and polymorphic state ⁴⁸ - and together with morphological data they give a coherent picture of a formulation's physicochemical properties and stability ⁴⁸⁻⁴⁹.

4. MECHANISMS OF CUTANEOUS DELIVERY AND SUITABILITY FOR INFANT SKIN

A topical formulation's clinical performance depends heavily on how the delivery system moves drug across the stratum corneum while controlling its distribution over time and depth. SLNs have drawn interest here^{50,57} because they promote dermal and transdermal penetration, act as a drug reservoir, maintain skin hydration and protect labile actives from degradation⁵⁸⁻⁵⁹ - properties that, in infantile dermatitis, map directly onto the therapeutic problem rather than being merely convenient. This section covers how nanoparticles interact with skin, the specific mechanisms by which SLNs improve delivery, and why those mechanisms matter for an immature barrier.

4.1 Skin penetration pathways for nanoparticles

The stratum corneum is the skin's principal barrier: corneocytes embedded in a continuous lipid matrix present substantial resistance to macromolecules and particulate carriers seeking deeper access⁶⁰. Nanoparticles cross this barrier via one of three routes - intercellular, transcellular or follicular (appendageal) - with the intercellular route generally dominant since tight junctions between corneocytes restrict most transcellular movement⁶¹⁻⁶². The relative contribution of each pathway shifts with the nanoparticle's physicochemical properties, its vehicle, skin condition, application site and local barrier integrity⁶³.

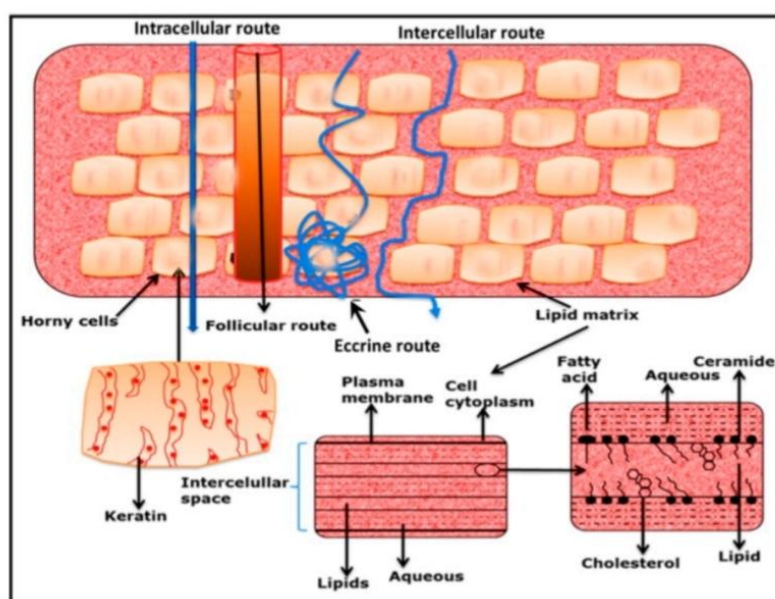


Figure 2. Routes of drug penetration through the skin, showing the intracellular, intercellular, follicular and eccrine (transappendageal) pathways across the horny cell layer and lipid matrix.

Via the **intercellular route**, drug molecules and small particles travel through the lipid-rich spaces between corneocytes, favouring small, lipophilic molecules that partition readily into that environment; the **transcellular route** requires passage through the corneocytes themselves and is considerably more resistant owing to their structural rigidity⁶²⁻⁶³. Most nanoparticle systems applied to skin accumulate in the outer stratum corneum rather than reaching transdermal depths, so it is prudent to assume intact particles do not generally penetrate far beyond the superficial layers⁶⁴. Hair follicles interrupt the stratum corneum at intervals, creating shunt pathways - the **follicular route** - that let nanoparticles bypass sections of the intact barrier and accumulate in the follicular infundibulum⁶⁵. This matters for pediatric use specifically: infant skin carries a higher density of hair follicles per unit area than adult skin, so follicular deposition likely contributes proportionally more to overall nanoparticle delivery in this age group⁶⁶.

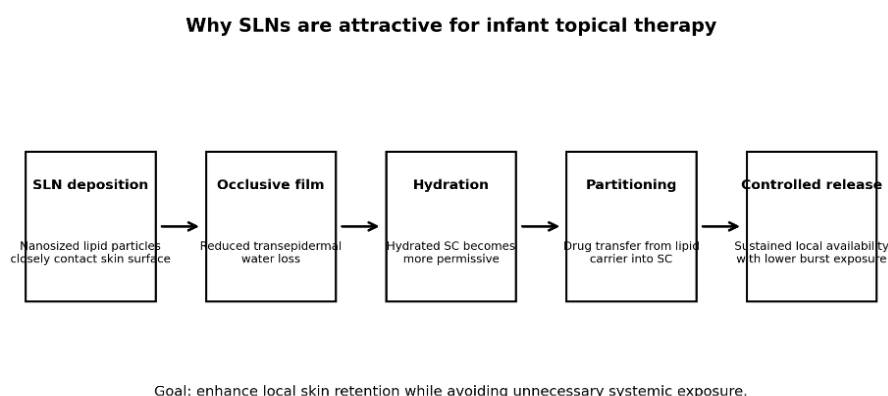
4.2 SLN-specific mechanisms of skin delivery

An SLN's performance as a topical vehicle depends on several interacting variables - particle size, lipid matrix type and crystalline structure, surfactant composition, drug distribution within the particle, formulation viscosity and the physiological state of the application site - which together shape three principal delivery mechanisms⁵⁸. On application, SLNs deposit on the skin surface and form a largely continuous **lipid film**, with the degree of occlusion depending on particle size, lipid concentration, dose and crystallinity⁶⁷⁻⁶⁸. By cutting transepidermal water loss, this film hydrates the outer stratum corneum, which loosens its lipids and widens intercorneocyte spaces, in turn helping the entrapped drug partition into the outer skin layers. The occlusive effect is therefore not

a cosmetic by-product but an active contributor to delivery⁶⁸ - and, in dermatitis, a therapeutic effect in its own right, since barrier hydration is itself a treatment goal.

Particle size gives SLNs a large surface-area-to-volume ratio and closer, more even contact with the irregular skin surface; smaller particles generally form better films and stronger occlusion, though smaller does not automatically mean deeper penetration. The available evidence indicates SLNs largely stay at the skin surface and accumulate in follicles, with deeper penetration depending heavily on formulation design^{56,64}. Finally, the solid lipid matrix acts as a **reservoir for sustained release**, retaining drug at the application site and releasing it gradually rather than in a burst - avoiding sharp systemic peaks and extending the local therapeutic effect⁵⁸, a behaviour of particular value in infants, where confining exposure to the treated area is a primary safety objective⁶⁹.

Figure 3. Proposed sequence linking SLN surface deposition, occlusion, hydration, drug partitioning and controlled local release in topical therapy for infant skin.



4.3 Relevance of these mechanisms to infant skin

As Section 1.2 outlined, infant skin differs structurally and functionally from adult skin: the stratum corneum is present from birth but continues maturing over the first few years, during which barrier and water-retention functions stay weaker than in adults⁶⁹. Infants also carry a substantially larger body surface area relative to body weight, exposing proportionally more of the body to any applied agent⁷⁰⁻⁷¹ - a combination that leaves infant skin more permeable and more prone to irritation and injury, and more so still with preterm birth, low birth weight or pre-existing dermatitis⁷¹.

This heightened permeability forces a balancing act: enough active agent reaching the affected skin against minimal systemic absorption. Conventional formulations, especially over large or damaged areas, carry a real risk of clinically significant systemic drug levels²⁰. SLNs' occlusive, slow-release behaviour addresses this directly, maintaining local drug availability while limiting the systemic fraction⁵⁸⁻⁵⁹ - the core argument for treating SLNs as a next-generation delivery platform here. It remains, though, a hypothesis grounded in mechanism rather than an established finding: dedicated pediatric pharmacokinetic and safety data are limited, and further work is needed before these benefits can be confirmed clinically⁶⁹.

A second advantage is biocompatibility - the lipid excipients used are typically physiological or biodegradable, and skin generally tolerates them well⁵⁹ - though biocompatibility is not set by the lipid core alone. Excipient purity, surfactant and preservative choice, particle size and shape, and applied dose all influence irritation potential, and these considerations carry extra weight for infant products, where careful excipient selection and formal irritation testing are mandatory development steps rather than optional ones⁴⁸.

4.4 Comparison with other nanocarriers used in dermatitis

SLNs are one of several nanocarrier classes under investigation for topical treatment of inflammatory and infectious skin disease. Liposomes and niosomes are vesicular carriers that can accommodate both hydrophilic and lipophilic drugs⁷²⁻⁷³, while polymeric nanoparticles rely on natural or synthetic polymers for controlled release and surface functionality⁷⁴. Table 4 reflects general trends from the literature rather than a strict ranking, since performance varies considerably with formulation and manufacturing method.

Table 4. Qualitative comparison of major nanocarrier classes relevant to topical dermatitis delivery.

Nanocarrier	Stability	Drug-loading capacity	Skin irritation potential	Relative cost
SLNs	Generally good; polymorphic transition possible	Good for lipophilic actives; matrix-dependent	Low with well-tolerated lipids; excipient choice still matters	Moderate
Liposomes	Lower; prone to oxidation, hydrolysis, leakage, vesicle fusion	Both hydrophilic and lipophilic drugs; retention design-dependent	Usually biocompatible; composition-dependent	Moderate to high
Niosomes	Often more storage-stable than liposomes	Hydrophilic and lipophilic drugs	Surfactant-dependent; excess surfactant may irritate	Low to moderate
Polymeric NPs	Often good; polymer- and process-dependent	Flexible; drug encapsulated or surface-associated	Depends on polymer chemistry and degradation products	Moderate to high

Compared with liposomes, SLNs have a solid, rigid matrix and do not depend on an intact phospholipid bilayer surviving storage, though liposomes retain a genuine edge in carrying aqueous- and lipid-soluble drugs in separate compartments⁷³. Niosomes swap phospholipids for non-ionic surfactants, giving good storage stability, simpler preparation and lower cost, though surfactant choice still governs both performance and tolerability^{75–76}. Polymeric nanoparticles offer real flexibility for controlled release and surface engineering, but their safety hinges on polymer chemistry, degradation by-products and residual processing materials, so toxicity or tissue-accumulation concerns must be judged formulation by formulation⁷⁴ - a formulation-specific uncertainty that is itself a disadvantage for an infant-facing product relative to a carrier built from physiological lipids.

4.5 Drug loading and interim synthesis

SLNs suit the poorly water-soluble actives relevant to dermatitis particularly well: lipophilic corticosteroids partition readily into the hydrophobic core⁷⁷, and calcineurin inhibitors (tacrolimus, pimecrolimus) and natural anti-inflammatory compounds (curcumin, quercetin) are similarly good candidates, though whether loading actually improves their anti-inflammatory efficiency at the target site remains an active research question⁷⁷. Successful loading depends on more than drug-lipid affinity - what matters practically is how much drug stays in the particle, the physical state it adopts, its compatibility with other components, its release kinetics and its eventual skin penetration⁴⁴. A highly crystalline matrix tends to expel drug over time, while a less ordered one accommodates a larger payload, which is the mechanistic basis for the NLC modification described in Section 3.2; DSC, XRD, drug-content analysis and in vitro release/permeation testing are the standard ways to confirm a formulation behaves as intended⁴⁸.

Taken as a whole, the case for SLNs in infantile dermatitis does not rest on nanoparticles crossing the skin barrier wholesale - it rests on their being engineered to interact with that barrier in a controlled, predictable way. Nanoscale size raises surface contact with the stratum corneum, the lipid matrix forms an occlusive film, the resulting hydration promotes drug partitioning into the superficial skin, and the solid matrix governs release rate. For infants, that combination could support effective local therapy while reducing the need for high concentrations of freely available drug⁵⁸ - but because infant skin behaves unpredictably with respect to permeability and clearance, any candidate formulation still needs formal evaluation for irritation, sensitization, local retention, permeation and systemic exposure, particularly where the barrier is already inflamed⁶⁹. SLNs remain a mechanistically sound platform for next-generation topical treatment here, especially for lipophilic drugs where the goal is sustained local presence with limited systemic exposure - but these are potential benefits awaiting pediatric-specific validation, not demonstrated clinical superiority.

5. EVIDENCE BASE: REPORTED SLN AND NLC FORMULATIONS FOR DERMATITIS-RELEVANT ACTIVES

5.1 Formulations across drug classes

Among corticosteroid systems, fluticasone propionate has been formulated as a semi-solid NLC using a quality-by-design approach, treating particle size and release rate as the critical attributes⁷⁸. The optimized formulation reached ~188 nm (PDI ~0.23) and released only about 11% of loaded drug over 24 hours - the slowest release

among the semi-solid NLCs tested, favouring once- or twice-daily dosing - and, unusually for this pool, was carried through to a rat model of chronic atopic dermatitis across skin-bleaching, paw-oedema, transepidermal-water-loss and histopathology endpoints, outperforming the conventional comparator on both efficacy and safety⁷⁸.

Tacrolimus poses a different problem: as a BCS class II molecule with low aqueous solubility and a molecular weight above 800 Da, it penetrates intact stratum corneum poorly, and the marketed ointment is widely reported to leave a greasy, tacky film - a compliance issue that matters more with an infant or toddler likely to rub the treated site⁷⁹. Chitosan-decorated tacrolimus SLNs prepared by solvent evaporation gave particles of 439-669 nm with 69-84% entrapment efficiency; zeta potential for the uncoated particles ran -26 to -15 mV but reversed to positive after chitosan coating, which the authors tie to better electrostatic interaction with the negatively charged skin surface and improved retention over inflamed sites⁷⁹.

For allergic contact dermatitis, where oral antihistamines are conventionally paired with topical steroids despite the atrophy risk, ebastine-loaded SLNs (Compritol® 888 ATO/Tween 80, optimized by central composite design) produced ~155 nm particles with zeta potential near -16 mV and entrapment above 78%⁸⁰. Embedded in a chitosan hydrogel - chitosan chosen partly for its own mild anti-inflammatory contribution - the system sustained release to 24 hours, somewhat slower in the hydrogel (~74%) than as a plain dispersion (~83%), and is one of the few studies here with a genuine disease-model endpoint: tested against picryl chloride-induced allergic contact dermatitis in BALB/c mouse ears over 15 days, ear swelling, mast cell counts and histopathology all showed symptom relief relative to untreated controls⁸⁰.

NSAID-loaded SLNs offer a useful reference point beyond dermatitis-specific actives. Etofenamate and ibuprofen were encapsulated in Compritol-based SLNs by fusion-emulsification, reaching entrapment efficiencies above 90%, particle sizes below 250 nm and twelve months of physical stability with scalable production⁸¹. Franz diffusion cell studies on human skin showed higher permeation than marketed comparator gels, and a rat paw-oedema model found SLN hydrogels containing roughly 8-fold less etofenamate and 16-fold less ibuprofen produced comparable anti-inflammatory effects to the commercial formulations, with no epidermal histological changes⁸¹ - arguably the single most persuasive *in vivo* evidence here that SLN encapsulation lowers the effective dose and, by extension, systemic exposure.

The plant-derived literature is smaller but growing. Curcumin and resveratrol, esterified with oleic acid and co-formulated with linolenic acid in an SLN system, showed no meaningful cytotoxicity in keratinocyte (NCTC 2544) or monocyte (THP-1) lines and suppressed IL-6 in the keratinocyte line - a cytokine directly implicated in dermatitis inflammation⁸². Quercetin has drawn more attention: an early solvent-free solid lipid nanosystem built on glyceryl dibehenate stayed physically stable for eight weeks refrigerated⁸³, and a more recent formulation embedding quercetin-loaded lipid nanoparticles in a Carbopol hydrogel validated a stability-indicating HPLC-PDA assay and confirmed retained antioxidant activity through accelerated and long-term storage⁸⁴. A related but non-SLN vesicular quercetin system tested on TPA-inflamed mouse dorsal skin found 5% PEG gave the greatest reduction in oedema and leukocyte infiltration among the ratios tested⁸⁵ - outside the strict SLN/NLC definition used elsewhere here, but useful supporting evidence for quercetin's topical anti-inflammatory activity once nanoencapsulated. Finally, leflunomide-loaded SLNs, developed for psoriasis rather than dermatitis, add further weight: prepared by microemulsion, the ellipsoidal particles (~273 nm, ~65% entrapment, >93% drug content) showed significantly improved photostability ($p < 0.001$) and better *in vitro* anti-inflammatory activity than free drug⁸⁶.

Across drug classes, a consistent picture holds: SLN/NLC encapsulation tends to improve stability and moderate the release profile relative to free drug, and in the studies that reached *in vivo* testing, this translated into comparable or better efficacy at meaningfully lower doses.

5.2 Permeation, *in vivo* evidence and the pediatric gap

Sustained rather than burst release is a recurring theme - most strikingly the fluticasone NLC's 11% release at 24 hours⁷⁸, with the ebastine and etofenamate/ibuprofen systems showing a comparable if less pronounced pattern⁸⁰⁻⁸¹ - and this favours less frequent dosing and lower cumulative skin exposure, a real consideration for infants whose barrier is still maturing. None of the permeation or retention data reviewed, however, was generated on infant or neonatal skin: every *ex vivo* and *in vitro* permeation study identified used adult human, porcine or synthetic membrane models, so extrapolating to an infant's thinner, more permeable stratum corneum remains an assumption rather than a finding - and since infant skin is more permeable, the likely direction of error is not a conservative one.

In vivo evidence is more substantial than a first pass through the abstracts suggests: three studies went beyond physicochemical characterization to a genuine disease or inflammation model - the fluticasone NLC in a rat chronic-AD model⁷⁸, the ebastine SLN hydrogel in a mouse allergic-contact-dermatitis model⁸⁰, and the etofenamate/ibuprofen SLN hydrogel in a rat paw-oedema model, which additionally showed meaningful dose-sparing⁸¹. The curcumin/resveratrol work stops at the cellular level⁸², and the TPA-induced mouse model used

for the vesicular quercetin system sits outside the strict SLN definition⁸⁵. Even on this stronger base, one gap holds across every study: not one tested a dermatitis formulation in a neonatal or juvenile animal model, let alone in infants - every in vivo data point traces back to adult humans or adult rodents. That gap, stated plainly rather than glossed over, is the largest single obstacle between the mechanistic case built in Section 4 and actual clinical use in infantile dermatitis.

5.3 Stability

Long-term stability was reported inconsistently across the pool, but the available results are encouraging: the etofenamate/ibuprofen SLNs held stable for twelve months⁸¹, the quercetin solid lipid nanosystem for eight weeks refrigerated⁸³, and the leflunomide system showed a statistically significant photostability gain over free drug⁸⁶. Where NLCs rather than conventional SLNs were used, studies did not report the drug-expulsion problems that affect first-generation SLNs on storage - consistent with the mechanistic rationale for NLC development given in Section 3.2.

5.4 Consolidated summary of reviewed formulations

Table 5. Summary of SLN and NLC formulations reported for dermatitis-relevant active pharmaceutical ingredients.

API	Carrier system	Particle size	Entrapment efficiency	Key finding	Ref.
Fluticasone propionate	QbD-optimized semi-solid NLC	~188 nm (PDI 0.229)	Not reported	Superior efficacy/safety vs. conventional formulation in rat chronic AD model	⁷⁸
Tacrolimus	Chitosan-coated SLN gel	439-669 nm	68.95-83.68%	Chitosan coating reverses zeta potential to positive; improved retention at inflamed sites	⁷⁹
Ebastine	SLN (Compritol/Tween 80) in chitosan hydrogel	155.2 +/- 1.5 nm	>78%	In vivo efficacy in mouse ACD model over 15 days; sustained 24 h release	⁸⁰
Etofenamate and ibuprofen	SLN (Compritol/Tween 80)	<250 nm	>90%	8-16-fold dose reduction with comparable efficacy in rat paw oedema model; 12-month stability	⁸¹
Curcumin + resveratrol	SLN (oleic acid-esterified, with linolenic acid)	Not reported	Not reported	No cytotoxicity in skin cell lines; IL-6 suppression	⁸²
Quercetin	Solvent-free solid lipid nanosystem (glyceryl dibehenate)	Not reported	Not reported	8-week refrigerated stability; optimized topical delivery of a poorly soluble flavonoid	⁸³
Quercetin	Lipid nanoparticle-Carbopol hydrogel	~104 nm (PDI 0.12)	Not reported	Validated stability-indicating HPLC-PDA assay; >90% retention and preserved antioxidant/anti-inflammatory activity after 180-day storage	⁸⁴
Leflunomide	SLN (microemulsion) in hydrogel	273.1 nm	65.25 +/- 0.95%	Improved photostability ($p < 0.001$) and in vitro anti-inflammatory activity vs. free drug	⁸⁶
Quercetin (vesicular, non-SLN)	PEG-containing phospholipid vesicles	Varies with PEG%	Not reported	Greatest oedema and leukocyte reduction at 5% PEG in TPA-induced mouse model	⁸⁵

6. SAFETY, REGULATORY STATUS AND TRANSLATIONAL OUTLOOK

6.1 Toxicological and biocompatibility profile

Most reports on cutaneous SLNs describe formulations built from glyceryl monostearate, stearic acid, Precirol® ATO 5, Compritol® 888 ATO, and mono-, di- and triglycerides, stabilized with Poloxamer 188 or 407, Tween 80, or lecithin⁸⁷⁻⁸⁸. Because these materials resemble native skin lipids and sebum, SLNs are generally considered low-irritancy and well tolerated, particularly against polymeric or strongly surfactant-based alternatives⁸⁸. Laboratory data offer qualified support: exposure of cultured HaCaT and NCTC 2544 keratinocytes and dermal fibroblasts to empty or drug-loaded SLNs did not reduce viability relative to controls, and topical application in animals produced no overt toxicity⁸⁸⁻⁸⁹. But a recent assessment of SLNs/NLCs for atopic dermatitis notes that long-term or continuous treatment, and nanoparticle accumulation or re-entry, have not been studied, despite reports that glyceryl monostearate can alter dermal fibroblast behaviour⁸⁸, and that nanoparticles have been found to persist in hair follicles for days after application⁸⁸ - both concerns that carry extra weight in children, whose follicular permeability is greater and whose smaller body size raises the systemic consequence of any absorbed dose.

The decisive limitation is the population studied. Nearly all work on SLN excipient tolerability, pharmaceutical performance and clinical behaviour has been done in adults or adult animals, and adult skin differs from infant skin in two material respects - barrier function and the ratio of skin surface area to body weight, the latter especially consequential since an equivalent applied dose has a much larger systemic effect in a child. None of the studies reviewed accounted for this, none examined SLN biocompatibility in infants or children, and the term “infantile dermatitis” appeared in the title or abstract of none of them. On the available evidence, SLNs have not been adequately characterized for this clinical setting and cannot presently be recommended for use in infants.

6.2 Regulatory considerations

ICH E11(R1), which governs pediatric clinical studies, holds that formulations should be chosen for a child's physiological needs rather than age alone, accounting for excipient safety, tolerability and the practicability of caregiver dosing⁹⁰. ICH E11A, on pediatric extrapolation, is directly relevant here given that virtually all available SLN safety and efficacy data comes from adult studies⁹¹. The FDA and EMA both align with ICH guidance and require pediatric formulations to be designed for ease of administration with demonstrably safe excipients⁹⁰⁻⁹², though regional divergence in requirements can still complicate simultaneous approval across jurisdictions. No SLN-based medicine for infantile dermatitis, atopic or otherwise, appears to have been approved by any major regulator; the only SLN-containing consumer product identified is a Polish moisturizer with no indication for inflammation or infection, and all reviewed AD studies were *in vitro*, in animals, or in adults. SLNs for cutaneous use in infantile dermatitis should accordingly be regarded as unapproved and unproven.

6.3 Patents, clinical trials and commercialization

A review of Google Patents and WIPO PATENTSCOPE, together with recent patent-focused literature, points to substantial research interest in lipid nanoparticle production and stabilization - patent WO201905525A1 describes a method for reducing Ostwald ripening⁹³, and patent reviews identify multiple filings covering dermal, ocular and injectable SLN systems⁹⁴⁻⁹⁵ - but no patent specifically addressing SLN production for infantile dermatitis was identified. A ClinicalTrials.gov search for SLN trials in pediatric skin conditions returned no relevant entries; the closest match is a terminated early-phase trial of a doxycycline nanoparticle hydrogel for atopic dermatitis (NCT02910011)⁹⁶, the only nanoparticle-medicine trial for AD identified in this review. A broader “atopic dermatitis” search returned roughly 76 trials, none concerning infantile dermatitis or lipid nanoparticles⁹⁷ - an absence of clinical data worth stating plainly rather than assuming away.

6.4 Manufacturing, scale-up and translational barriers

Laboratory-scale SLN preparation by high-shear homogenization, hot/cold homogenization or ultrasonication does not translate directly to continuous, high-volume production at therapeutic quality and consistency. Scale-up reviews point to high-pressure homogenization and membrane-based methods as the most promising routes, though both need careful optimization of temperature, pressure and flow rate^{53,98-99}, and solvent removal, where required, adds further process complexity. Continuous manufacturing is generally disfavoured in pharmaceutical production because of batch-qualification difficulty, though it has worked elsewhere - notably for complex lipid nanoparticles in mRNA vaccine delivery - so a comparable approach for SLNs is plausible but would need substantial development work.

A further challenge is the metastability of SLN dispersions: Ostwald ripening, where the lipid shifts from a less to a more stable crystal form with effective dissolution of smaller particles, degrades the system over time¹⁰⁰⁻¹⁰¹, and particles may also agglomerate, raising mean size and suspension viscosity. Both phenomena matter more, not less, for a product meant for frequent application to a child, which must hold specification across a

commercially viable shelf life. Cost is the final factor: a formulation with these properties - including any cold-chain requirement - combined with the extra preclinical and clinical work needed for pediatric safety and efficacy, would run considerably more expensive than a conventional cream or gel with the same active, and whether that premium is justified depends on the indication and the size of the advantage SLN delivery actually provides.

6.5 Future perspectives

Several studies have combined lipid nanoparticles with secondary delivery vehicles - SLNs embedded in chitosan hydrogels, Carbopol gels and oleogels^{80,102-103} - improving spreadability, application comfort and skin residence time, a meaningful consideration for sensitive, easily irritated pediatric skin where site retention drives therapeutic effect. The hydrogel also appears to benefit the nanoparticles themselves, restricting particle mobility and thereby reducing aggregation and Ostwald ripening. Among the options reviewed, the SLN-hydrogel combination looks the most promising basis for a pediatric product.

Because the reviewed literature does not address SLN use in children, pharmacokinetic, safety and efficacy studies in this population are the clear priority. Given the range of physiology across developmental stages, a single SLN formulation is unlikely to suit the whole pediatric age range - percutaneous permeability, surface area and body weight all vary considerably from neonate to adolescent, each affecting a delivery vehicle's safety and efficacy. A more defensible strategy would stratify by age and optimize composition - lipid selection, particle size, applied quantity - separately for each group rather than applying one formulation across all patients, consistent with ICH E11(R1)'s emphasis on physiological differences in pediatric study design⁹⁰⁻⁹¹.

7. CONCLUSION

Solid lipid nanoparticles form a mechanistically coherent, well-tolerated platform with genuine potential as a topical vehicle for infantile dermatitis. The argument holds together internally: infant skin is thinner, more permeable and disproportionately large relative to body mass, which makes systemic absorption the principal safety constraint on topical therapy, and SLNs answer that constraint directly - forming an occlusive film that hydrates an already-compromised barrier, depositing preferentially in the stratum corneum and follicles rather than penetrating transdermally, and releasing drug slowly from a solid matrix. The formulation literature backs these claims: encapsulation consistently improved stability and moderated release, and in the studies that reached animal disease models it matched or beat free-drug efficacy at substantially lower doses.

What the literature does not support is the final step. No study identified tested an SLN or NLC dermatitis formulation on infant or neonatal skin, in a neonatal or juvenile animal model, or in infants themselves; no clinical trial of such a formulation exists; no regulator has approved one; no patent addresses the indication. The biocompatibility data, while reassuring, come entirely from adult cells, adult animals and adult patients, and say nothing about long-term exposure, nanoparticle accumulation, or the consequences of prolonged follicular retention in a population with greater follicular permeability. Translational obstacles compound this gap: scale-up needs process development that has not been undertaken for this application, Ostwald ripening and agglomeration threaten the shelf life a pediatric product would require, and the cost premium over a conventional cream is substantial.

The honest position sits between the two extremes. SLN-based delivery for infantile dermatitis is not an established therapy and should not be described as one, but it is not an unfounded proposition either, since the mechanism is sound and the adult evidence points consistently in its favour. It is a well-supported hypothesis still waiting on the pediatric-specific pharmacokinetic, irritation and safety data that would let it be tested - and generating that data, starting with juvenile animal models and age-stratified formulation optimization, is the necessary next step. Defining precisely which evidence is missing is, in the meantime, the most useful contribution a review like this one can make.

Acknowledgements

The authors thank the Department of Pharmaceutics, Chemists College of Pharmaceutical Sciences and Research, Varikoli, Ernakulam, for institutional support in the preparation of this review. The authors declare no conflicts of interest and received no external funding for this work.

REFERENCES

1. Avena-Woods C. Overview of atopic dermatitis. *Am J Manag Care*. 2017;23(8 Suppl):S115-23.
2. Werfel T, Heratizadeh A, Aberer W, Ahrens F, Augustin M, Biedermann T, et al. S2k guideline on diagnosis and treatment of atopic dermatitis — short version. *Allergo J Int*. 2016;25:82-95.
3. Dhar S, Kanwar AJ. Epidemiology and clinical pattern of atopic dermatitis in a North Indian pediatric population. *Pediatr Dermatol*. 1998;15:347-51.
4. Sarkar R, Kanwar AJ. Clinico-epidemiological profile and factors affecting severity of atopic dermatitis in North Indian children. *Indian J Dermatol*. 2004;49:117-22.

5. Dhar S, Mandal B, Ghosh A. Epidemiology and clinical pattern of atopic dermatitis in 100 children seen in a city hospital. *Indian J Dermatol.* 2002;47:202–4.
6. Stuetz A, Baumann K, Grassberger M, Wolff K, Meingassner JG. Discovery of topical calcineurin inhibitors and pharmacological profile of pimecrolimus. *Int Arch Allergy Immunol.* 2006;141(3):199–212. DOI: 10.1159/000095289.
7. Aschoff R, Schmitt J, Knuschke P, Koch E, Bräutigam M, Meurer M. Evaluation of the atrophogenic potential of hydrocortisone 1% cream and pimecrolimus 1% cream in uninvolved forehead skin of patients with atopic dermatitis using optical coherence tomography. *Exp Dermatol.* 2011;20(10):832–6. DOI: 10.1111/j.1600-0625.2011.01335.x.
8. Li AW, Yin ES, Antaya RJ. Topical corticosteroid phobia in atopic dermatitis: a systematic review. *JAMA Dermatol.* 2017;153(10):1036–42. DOI: 10.1001/jamadermatol.2017.2437.
9. Siegfried EC, Jaworski JC, Hebert AA. Topical calcineurin inhibitors and lymphoma risk: evidence update with implications for daily practice. *Am J Clin Dermatol.* 2013;14(3):163–78. DOI: 10.1007/s40257-013-0020-1.
10. Stamatas GN, Nikolovski J, Mack MC, Kollias N. Infant skin physiology and development during the first years of life: a review of recent findings based on in vivo studies. *Int J Cosmet Sci.* 2011;33:17–24.
11. Schmid-Wendtner MH, Korting HC. The pH of the skin surface and its impact on the barrier function. *Skin Pharmacol Physiol.* 2006;19:296–302.
12. Vitellaro-Zuccarello L, Cappelletti S, Dal Pozzo Rossi V, Sari-Gorla M. Stereological analysis of collagen and elastic fibers in the normal human dermis: variability with age, sex, and body region. *Anat Rec.* 1994;238:153–62.
13. Nikolovski J, Stamatas GN, Kollias N, Wiegand BC. Barrier function and water-holding and transport properties of infant stratum corneum are different from adult and continue to develop through the first year of life. *J Invest Dermatol.* 2008;128(7):1728–36. DOI: 10.1038/sj.jid.5701239.
14. Silverman RA. Pediatric dermatology. In: Schachner LA, Hansen RC, editors. 4th ed. Philadelphia (PA): Elsevier; 2014.
15. Oranges T, Dini V, Romanelli M. Skin physiology of the neonate and infant: clinical implications. *Adv Wound Care (New Rochelle).* 2015;4:587–95.
16. Luger TA, Lahfa M, Fölster-Holst R, Gulliver WP, Allen R, Molloy S, et al. Long-term safety and tolerability of pimecrolimus cream 1% and topical corticosteroids in adults with moderate to severe atopic dermatitis. *J Dermatolog Treat.* 2004;15:169–78.
17. Peserico A, Städtler G, Sebastian M, Fernandez RS, Vick K, Bieber T. Reduction of relapses of atopic dermatitis with methylprednisolone aceponate cream twice weekly in addition to maintenance treatment with emollient: a multicentre, randomized, double-blind, controlled study. *Br J Dermatol.* 2008;158:801–7.
18. Fukuie T, Hirakawa S, Narita M, Nomura I, Matsumoto K, Tokura Y, et al. Potential preventive effects of proactive therapy on sensitization in moderate to severe childhood atopic dermatitis: a randomized, investigator-blinded, controlled study. *J Dermatol.* 2016;43:1283–92.
19. British Medical Association, Royal Pharmaceutical Society of Great Britain. British National Formulary. 69th ed. London: BMA and RPS;
20. Hong E, Smith S, Fischer G. Evaluation of the atrophogenic potential of topical corticosteroids in pediatric dermatology patients. *Pediatr Dermatol.* 2011;28:393–6.
21. Sigurgeirsson B, Boznanski A, Todd G, Vertruyen A, Schuttelaar ML, Zhu X, et al. Safety and efficacy of pimecrolimus in atopic dermatitis: a 5-year randomized trial. *Pediatrics.* 2015;135:597–606.
22. Ashcroft DM, Dimmock P, Garside R, Stein K, Williams HC. Efficacy and tolerability of topical pimecrolimus and tacrolimus in the treatment of atopic dermatitis: meta-analysis of randomized controlled trials. *BMJ.* 2005;330:516.
23. Ring J, Alomar A, Bieber T, Deleuran M, Fink-Wagner A, Gelmetti C, et al. Guidelines for treatment of atopic eczema (atopic dermatitis) part II. *J Eur Acad Dermatol Venereol.* 2012;26:1176–93.
24. Siegfried EC, Jaworski JC, Kaiser JD, Hebert AA. Systematic review of published trials: long-term safety of topical corticosteroids and topical calcineurin inhibitors in pediatric patients with atopic dermatitis. *BMC Pediatr.* 2016;16:75.
25. Nakamura M, Uemura K, Nemoto O, Miyachi Y. Evaluation of an optimal method for topical application of moisturizer. *Skin Res.* 2006;5:311.
26. Grimalt R, Mengeaud V, Cambazard F; Study Investigators' Group. The steroid-sparing effect of an emollient therapy in infants with atopic dermatitis: a randomized controlled study. *Dermatology.* 2007;214:61–7.
27. Hon KL, Ching GK, Leung TF, Choi CY, Lee KK, Ng PC. Estimating emollient usage in patients with eczema. *Clin Exp Dermatol.* 2010;35:22–6.
28. Draelos ZD. An evaluation of prescription device moisturizers. *J Cosmet Dermatol.* 2009;8:40–3.

29. Panahi Y, Davoudi SM, Sahebkar A, Beiraghdar F, Dadjo Y, Feizi I, et al. Efficacy of Aloe vera/olive oil cream versus betamethasone cream for chronic skin lesions following sulfur mustard exposure: a randomized double-blind clinical trial. *Cutan Ocul Toxicol.* 2012;31:95–103.
30. Intahphuak S, Khonsung P, Panthong A. Anti-inflammatory, analgesic and antipyretic activities of virgin coconut oil. *Pharm Biol.* 2010;48(2):151–7. DOI: 10.3109/13880200903062614.
31. Akihisa T, Kojima N, Kikuchi T, Yasukawa K, Tokuda H, Masters ET, et al. Anti-inflammatory and chemopreventive effects of triterpene cinnamates and acetates from shea fat. *J Oleo Sci.* 2010;59:273–80.
32. Varothai S, Nitayavardhana S, Kulthanan K. Moisturizers for patients with atopic dermatitis. *Asian Pac J Allergy Immunol.* 2013;31:91.
33. Richardson M. Understanding the structure and function of the skin. *Nurs Times.* 2003;99:46–8.
34. Bouwstra JA, Ponc M. The skin barrier in healthy and diseased state. *Biochim Biophys Acta.* 2006;1758:2080–95.
35. de Souza Guedes L, Martinez RM, Bou-Chacra NA, Velasco MVR, Rosado C, Baby AR. An overview on topical administration of carotenoids and coenzyme Q10 loaded in lipid nanoparticles. *Antioxidants (Basel).* 2021;10:1034.
36. Fore J. A review of skin and the effects of aging on skin structure and function. *Ostomy Wound Manage.* 2006;52:24–35.
37. Bouwstra J. Structure of the skin barrier and its modulation by vesicular formulations. *Prog Lipid Res.* 2003;42:1–36.
38. Bukhari SI, Imam SS, Ahmad MZ, Vuddanda PR, Alshehri S, Mahdi WA, et al. Recent progress in lipid nanoparticles for cancer theranostics: opportunity and challenges. *Pharmaceutics.* 2021;13:840.
39. Garg A, Sharma GS, Goyal AK, Ghosh G, Si SC, Rath G. Recent advances in topical carriers of anti-fungal agents. *Heliyon.* 2020;6:e04663.
40. DeLouise LA. Applications of nanotechnology in dermatology. *J Invest Dermatol.* 2012;132:964–75.
41. Lason E. Topical administration of terpenes encapsulated in nanostructured lipid-based systems. *Molecules.* 2020;25:5758.
42. Battaglia L, Gallarate M. Lipid nanoparticles: state of the art, new preparation methods and challenges in drug delivery. *Expert Opin Drug Deliv.* 2012;9:497–508.
43. Nyström AM, Fadeel B. Safety assessment of nanomaterials: implications for nanomedicine. *J Control Release.* 2012;161:403–8.
44. Viegas C, Patrício AB, Prata JM, Nadhman A, Chintamaneni PK, Fonte P. Solid lipid nanoparticles vs. nanostructured lipid carriers: a comparative review. *Pharmaceutics.* 2023;15(6):1593. DOI: 10.3390/pharmaceutics15061593.
45. Munir M, Zaman M, Waqar MA, Khan MA, Alvi MN. Solid lipid nanoparticles: a versatile approach for controlled release and targeted drug delivery. *J Liposome Res.* 2024;34(2):335–48. DOI: 10.1080/08982104.2023.2268711.
46. Gao S, McClements DJ. Formation and stability of solid lipid nanoparticles fabricated using phase inversion temperature method. *Colloids Surf A Physicochem Eng Asp.* 2016;499:79–87. DOI: 10.1016/j.colsurfa.2016.03.065.
47. Iqbal MA, Md S, Sahni JK, Baboota S, Dang S, Ali J. Nanostructured lipid carriers system: recent advances in drug delivery. *J Drug Target.* 2012;20(10):813–30. DOI: 10.3109/1061186X.2012.716845.
48. Severino P, Andreani T, Macedo AS, Fangueiro JF, Santana MHA, Silva AM, et al. Current state-of-art and new trends on lipid nanoparticles (SLN and NLC) for oral drug delivery. *J Drug Deliv.* 2012;2012:750891. DOI: 10.1155/2012/750891.
49. Wei CC, Ge ZQ. Influence of electrolyte and poloxamer 188 on the aggregation kinetics of solid lipid nanoparticles (SLNs). *Drug Dev Ind Pharm.* 2012;38(9):1084–9. DOI: 10.3109/03639045.2011.640331.
50. Müller RH, Radtke M, Wissing SA. Solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) in cosmetic and dermatological preparations. *Adv Drug Deliv Rev.* 2002;54(Suppl 1):S131–55. DOI: 10.1016/S0169-409X(02)00118-7.
51. Stefanov S, Andonova V, Peneva P, Kassarova M. Technological strategies for the preparation of lipid nanoparticles: an updated review. *Pharmacia.* 2023;70(3):449–63. DOI: 10.3897/pharmacia.70.e108119.
52. Subroto E, Andoyo R, Indarto R. Solid lipid nanoparticles: review of the current research on encapsulation and delivery systems for active and antioxidant compounds. *Antioxidants (Basel).* 2023;12(3):633. DOI: 10.3390/antiox12030633.
53. Khairnar SV, Pagare P, Thakre A, Nambiar AR, Junnuthula V, Abraham MC, et al. Review on the scale-up methods for the preparation of solid lipid nanoparticles. *Pharmaceutics.* 2022;14(9):1886. DOI: 10.3390/pharmaceutics14091886.
54. Mehnert W, Mäder K. Solid lipid nanoparticles: production, characterization and applications. *Adv Drug Deliv Rev.* 2001;47(2–3):165–96. DOI: 10.1016/S0169-409X(01)00105-3.

56. Müller RH, Mäder K, Gohla S. Solid lipid nanoparticles (SLN) for controlled drug delivery — a review of the state of the art. *Eur J Pharm Biopharm.* 2000;50(1):161–77. DOI: 10.1016/S0939-6411(00)00087-4.
57. Mardhiah Adib Z, Ghanbarzadeh S, Kouhsoltani M, Yari Khosroshahi A, Hamishehkar H. The effect of particle size on the deposition of solid lipid nanoparticles in different skin layers: a histological study. *Adv Pharm Bull.* 2016;6(1):31–6. DOI: 10.15171/apb.2016.06.
58. Souto EB, Müller RH. Cosmetic features and applications of lipid nanoparticles (SLN, NLC). *Int J Cosmet Sci.* 2008;30(3):157–65. DOI: 10.1111/j.1468-2494.2008.00433.x.
59. Liu M, Wen J, Sharma M. Solid lipid nanoparticles for topical drug delivery: mechanisms, dosage form perspectives, and translational status. *Curr Pharm Des.* 2020;26(27):3203–17. DOI: 10.2174/1381612826666200526145706.
60. Pardeike J, Hommoss A, Müller RH. Lipid nanoparticles (SLN, NLC) in cosmetic and pharmaceutical dermal products. *Int J Pharm.* 2009;366(1–2):170–84. DOI: 10.1016/j.ijpharm.2008.10.003.
61. Almeida AJ, Souto E. Solid lipid nanoparticles as a drug delivery system for peptides and proteins. *Adv Drug Deliv Rev.* 2007;59(6):478–90. DOI: 10.1016/j.addr.2007.04.007.
62. Prausnitz MR, Langer R. Transdermal drug delivery. *Nat Biotechnol.* 2008;26(11):1261–8. DOI: 10.1038/nbt.1504.
63. Hadgraft J, Lane ME. Skin permeation: the years of enlightenment. *Int J Pharm.* 2005;305(1–2):2–12. DOI: 10.1016/j.ijpharm.2005.07.014.
64. Barry BW. Novel mechanisms and devices to enable successful transdermal drug delivery. *Eur J Pharm Sci.* 2001;14(2):101–14. DOI: 10.1016/S0928-0987(01)00167-1.
65. Wissing SA, Müller RH. Solid lipid nanoparticles as carrier for sunscreens: in vitro release and in vivo skin penetration. *J Control Release.* 2002;81(3):225–33. DOI: 10.1016/S0168-3659(02)00056-1.
66. Vogt A, Mandt N, Lademann J, Schaefer H, Blume-Peytavi U. Follicular targeting: distribution of fluorescent dyes in the hair follicles of human scalp skin. *Eur J Pharm Biopharm.* 2006;64(2):223–30.
67. Stamatias GN, Nikolovski J, Luedtke MA, Kollias N, Wiegand BC. Infant skin microstructure assessed in vivo differs from adult skin in organization and at the cellular level. *Pediatr Dermatol.* 2010;27(2):125–31. DOI: 10.1111/j.1525-1470.2009.00973.x.
68. Wissing SA, Müller RH. The influence of the crystallinity of lipid nanoparticles on their occlusive properties. *Int J Pharm.* 2002;242(1–2):377–9. DOI: 10.1016/S0378-5173(02)00220-X.
69. Üner M, Wissing SA, Yener G, Müller RH. Skin moisturizing effect and skin penetration of ascorbyl palmitate entrapped in solid lipid nanoparticles and nanostructured lipid carriers incorporated into hydrogel. *Pharmazie.* 2005;60(10):751–5.
70. Telofski LS, Morello AP 3rd, Mack Correa MC, Stamatias GN. The infant skin barrier: can we preserve, protect, and enhance the barrier? *Dermatol Res Pract.* 2012;2012:198789. DOI: 10.1155/2012/198789.
71. Chiou YB, Blume-Peytavi U. Stratum corneum maturation: a review of neonatal skin function. *Skin Pharmacol Physiol.* 2004;17(2):57–66. DOI: 10.1159/000076015.
72. Kalia YN, Nonato LB, Lund CH, Guy RH. Development of skin barrier function in premature infants. *J Invest Dermatol.* 1998;111(2):320–6. DOI: 10.1046/j.1523-1747.1998.00289.x.
73. Choi MJ, Maibach HI. Liposomes and niosomes as topical drug delivery systems. *Skin Pharmacol Physiol.* 2005;18(5):209–19. DOI: 10.1159/000086666.
74. Akbarzadeh A, Rezaei-Sadabady R, Davaran S, Joo SW, Zarghami N, Hanifehpour Y, et al. Liposome: classification, preparation, and applications. *Nanoscale Res Lett.* 2013;8(1):102. DOI: 10.1186/1556-276X-8-102.
75. Zielińska A, Carreiró F, Oliveira AM, Neves A, Pires B, Venkatesh DN, et al. Polymeric nanoparticles: production, characterization, toxicology and ecotoxicology. *Molecules.* 2020;25(16):3731. DOI: 10.3390/molecules25163731.
76. Moghassemi S, Hadjizadeh A. Nano-niosomes as nanoscale drug delivery systems: an illustrated review. *J Control Release.* 2014;185:22–36. DOI: 10.1016/j.jconrel.2014.04.015.
77. Abu-Huwaij R, Alkarawi A, Salman D, Alkarawi F. Exploring the use of niosomes in cosmetics for efficient dermal drug delivery. *Pharm Dev Technol.* 2023;28(7):708–18. DOI: 10.1080/10837450.2023.2233613.
78. Pham DTT, Tran PHL, Tran TTD. Development of solid dispersion lipid nanoparticles for improving skin delivery. *Saudi Pharm J.* 2019;27(7):1019–24. DOI: 10.1016/j.jsps.2019.08.004.
79. Amasya G, Badilli U, Sengel-Turk CT, Onay-Besikci A, Aksu B, Tarimci N. Development and in vivo evaluation of a novel semi-solid nanostructured lipid carrier of fluticasone propionate for the treatment of atopic dermatitis using a quality by design approach. *J Drug Deliv Sci Technol.* 2025;114:107491.
80. Khan AS, Shah KU, Al Mohaini M, Alsalman AJ, Al Hawaj MA, Alhashem YN, et al. Tacrolimus-loaded solid lipid nanoparticle gel: formulation development and in vitro assessment for topical applications. *Gels.* 2022;8(2):129. DOI: 10.3390/gels8020129.

83. Kazim T, Tariq A, Usman M, Ayoob MF, Khan A. Chitosan hydrogel for topical delivery of ebastine loaded solid lipid nanoparticles for alleviation of allergic contact dermatitis. *RSC Adv.* 2021;11(59):37413–25. DOI: 10.1039/D1RA06283B.
84. Mancini G, Gonçalves LMD, Marto J, Carvalho FA, Simões S, Ribeiro HM, et al. Increased therapeutic efficacy of SLN containing etofenamate and ibuprofen in topical treatment of inflammation. *Pharmaceutics.* 2021;13(3):328. DOI: 10.3390/pharmaceutics13030328.
85. Cassano R, Serini S, Curcio F, Trombino S, Calviello G. Preparation and study of solid lipid nanoparticles based on curcumin, resveratrol and capsaicin containing linolenic acid. *Pharmaceutics.* 2022;14(8):1593. DOI: 10.3390/pharmaceutics14081593.
86. Bose S, Du Y, Takhistov P, Michniak-Kohn B. Formulation optimization and topical delivery of quercetin from solid lipid based nanosystems. *Int J Pharm.* 2013;441(1–2):56–66. DOI: 10.1016/j.ijpharm.2012.12.013.
87. Tangjidadapichai C, Tadtong S, Chittasupho C, Samee W. Preparation of quercetin-loaded lipid nanoparticle-embedded hydrogels and stability studies. *Molecules.* 2026;31(14):2539. DOI: 10.3390/molecules31142539.
88. Caddeo C, Díez-Sales O, Pons R, Fernández-Busquets X, Fadda AM, Manconi M. Topical anti-inflammatory potential of quercetin in lipid-based nanosystems: in vivo and in vitro evaluation. *Pharm Res.* 2014;31(4):959–68. DOI: 10.1007/s11095-013-1215-0.
89. Alhelal HM, Mehta S, Kadian V, Kakkar V, Tanwar H, Rao R, et al. Solid lipid nanoparticles embedded hydrogels as a promising carrier for retarding irritation of leflunomide. *Gels.* 2023;9(7):576. DOI: 10.3390/gels9070576.
90. Lazov C, Yoncheva K, Slavkova M. The significance of solid lipid nanoparticles (SLNs) and nanostructured lipid carriers (NLCs) in the treatment of atopic dermatitis. *Sci Pharm.* 2026;94(1):19. DOI: 10.3390/scipharm94010019.
91. Souto EB, Baldim I, Oliveira WP, Rao R, Yadav N, Gama FM, et al. SLN and NLC for topical, dermal, and transdermal drug delivery. *Expert Opin Drug Deliv.* 2020;17(3):357–77. DOI: 10.1080/17425247.2020.1727883.
92. Siddique MI, Katas H, Amin MCI, Ng SF, Zulfakar MH, Jamil A. In vivo dermal pharmacokinetics, efficacy, and safety of skin targeting nanoparticles for corticosteroid treatment of atopic dermatitis. *Int J Pharm.* 2016;507(1–2):72–82. DOI: 10.1016/j.ijpharm.2016.05.005.
93. European Medicines Agency. ICH E11(R1) guideline on clinical investigation of medicinal products in the pediatric population: addendum. Amsterdam: EMA; 2017. EMA/CPMP/ICH/2711/1999.
94. US Food and Drug Administration. E11A pediatric extrapolation: guidance for industry. Silver Spring (MD): FDA; 2024.
95. Zisowsky J, Krause A, Dingemans J. Drug development for pediatric populations: regulatory aspects. *Pharmaceutics.* 2010;2(4):364–88. DOI: 10.3390/pharmaceutics2040364.
96. Solid nanoparticle formulation of water insoluble pharmaceutical substances with reduced Ostwald ripening. WO patent WO201905525A1. 2019 Mar 21.
97. Khames A, Khaleel MA, El-Badawy MF, El-Nezhawy AOH. Natamycin solid lipid nanoparticles — sustained ocular delivery system of higher corneal penetration against deep fungal keratitis: preparation and optimization. *Int J Nanomedicine.* 2019;14:2515–31. DOI: 10.2147/IJN.S190502.
98. Paliwal R, Paliwal SR, Kenwat R, Kurmi BD, Sahu MK. Solid lipid nanoparticles: a review on recent perspectives and patents. *Expert Opin Ther Pat.* 2020;30(3):179–94. DOI: 10.1080/13543776.2020.1720649.
99. ClinicalTrials.gov. Topical NanoDox for atopic dermatitis. Bethesda (MD): National Library of Medicine. Identifier NCT02910011.
100. Waligóra-Dziwak K, Dańczak-Pazdrowska A, Jenerowicz D. A comprehensive review of biologics in phase III and IV clinical trials for atopic dermatitis. *J Clin Med.* 2024;13(14):4001. DOI: 10.3390/jcm13144001.
101. Muthu MS, Wilson B. Challenges posed by the scale-up of nanomedicines. *Nanomedicine (Lond).* 2012;7(3):307–9. DOI: 10.2217/nnm.12.3.
102. Mehta M, Bui TA, Yang X, Aksoy Y, Goldys EM, Deng W. Lipid-based nanoparticles for drug/gene delivery: an overview of the production techniques and difficulties encountered in their industrial development. *ACS Mater Au.* 2023;3(6):600–19. DOI: 10.1021/acsmaterialsau.3c00032.
103. Attama AA, Momoh MA, Builders PF. Lipid nanoparticulate drug delivery systems: a revolution in dosage form design and development. In: Fikai A, Grumezescu AM, editors. *Nanostructures for oral medicine.* London: IntechOpen; 2012. DOI: 10.5772/50486.
104. Nguyen VH, Thuy VN, Van TV, Dao AH, Lee BJ. Nanostructured lipid carriers and their potential applications for versatile drug delivery via oral administration. *OpenNano.* 2022;8:100064. DOI: 10.1016/j.onano.2022.100064.
- 105.

112. Malik S, Rehman AU, Khan MFA, Ahmed N, Al Nasir MH. Solid lipid nanoparticles based topical hydrogel for potential treatment of infantile hemangioma. *BioNanoScience*. 2024;14(3):2964–75. DOI: 10.1007/s12668-024-01569-9.
113. 10.1007/s12668-024-01569-9.
114. Ebrahimian K, Dabbagh Moghaddam F, Tavangari SM. Next-generation atopic dermatitis treatments based on nanotechnology as an effective and low-side-effect solution. *Oxford Open Mater Sci*. 2026;6(1):itag003. DOI: 10.1093/oxfmat/itag003